

Regiospecific Benzoylation of Electron-Deficient N-Heterocycles with Methylbenzenes via a Minisci-Type Reaction

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Supporting Information

ABSTRACT: A regioselective cross-dehydrogenative coupling between electron-deficient N-heterocycles (isoquinoline, quinolines, and quinoxalines) and methylbenzenes leading to regiospecific C-aroylation has been accomplished using AlCl₃ as the catalyst in the presence of oxidant TBHP. This protocol is a practical alternative to the classical Minisci reaction.



■ INTRODUCTION

In organic chemistry, one of the most important and fundamental challenge is to build C-C bonds in a rapid and efficient manner. Of significant interest are methods that provide access to molecules in step- and atom-economic fashion from readily available precursors. In the past few decades, the construction of C-C bonds through C-H bond activation is a rapidly expanding field of research as it provides an atom-economical and shorter route for the synthesis of organic compounds and offers substantial benefits.² In this context, cross-dehydrogenative coupling between two different C-H bonds represent an useful alternative approach toward C-C bond formation between organic components through the functionalization of all types (sp, sp², sp³) of C-H bonds. 1a,d,e,3 Mild reactivity and poor site selective are the two important challenges that need to be addressed in CDC protocols. Hence, the development of new and efficient CDC reactions in which such challenges can be overcome are of great significance in synthetic organic chemistry.

Nitrogenous heterocycles are widely distributed in nature and present in large proportion in commercial drugs.⁴ These heterocycles also have enormous applications in both chemistry and biology. 5 Heterocyclic moieties bearing an acyl group have been found in drugs which are important in pharmacological studies. 4 Until now, a number of methods have been developed for the synthesis of electron-deficient heterocycles, but their functionalization using cross-dehydrogenative coupling is far less visited. In contrast to electron-rich heterocycles, 6a-g acylation of electron-deficient heterocycles is much more challenging, and only a few reports are available. 6h-j Among these, the Minisci reaction is the most commonly used approach, 4a,6i,8-11 which involves the addition of an in situ generated nucleophilic acyl radical from aldehyde to an electron-deficient heterocycle, although it represents a straightforward strategy but suffers from certain drawbacks such as harsh reaction conditions, poor site selectivity, limited substrate scope, and the use of transition-metal salts up to stoichiometric amounts. 8a To overcome these aspects, the Antonchick^{6h} and Prabhu¹² groups independently reported

metal-free analogues of the Minisci reaction under ambient conditions (Scheme 1, known methods).

Ref 6h

Scheme 1. Reported and Designed Route for the Direct Acylation

Recently, our group has developed a number of CDC protocols for the construction of C–C and C–O bonds using alkylbenzenes as the surrogates of $ArCH_2O$ –, 13a ArCO–, $^{13b-d}$ $ArCH_2$ –, 13e and ArCOO– under oxidative conditions. In continuation to our efforts in utilizing alkylbenzenes as different surrogates via metal and metal-free C-H functionalization strategies, we envisaged that an acyl radical (generated in situ from methylarenes under oxidative conditions) could be utilized for the direct acylation of N-heterocycles. Antonchick et al.6h in their report demonstrated that the in situ generated CF₃COOH from PhI(OCOCF₃)₂ protonates the N-atom of heterocycle, therefore making α -C to nitrogen more electrophilic in nature. This facilitates the attack of the nucleophilic acyl radical on to the α -C. Intrigued by the key mechanistic

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features, we anticipated that the direct use of CF_3COOH could generate a similar protonated intermediate. On the other hand, employing TBHP as the oxidant is expected to afford the other coupling counterpart to the acyl radical from methylarenes.

RESULTS AND DISCUSSION

To give a practical shape to this coupling concept, a reaction between isoquinoline (1) and p-xylene (d) was initiated in the presence of CF₃COOH and TBHP at 110 °C. As hypothesized, the reaction provided C1-aroylated product (1d) but in a low yield of 26% (Table 1, entry 1). In a quest to improve the yield,

Table 1. Screening of Reaction Conditions^a

entry	catalyst (mol %)	oxidant (equiv)	temp ($^{\circ}$ C)	$yield^b$ (%)
1	CF ₃ COOH (100)	TBHP (3)	110	26
2	CH ₃ COOH (100)	TBHP (3)	110	09
3	CF ₃ SO ₃ H (100)	TBHP (3)	110	13
4	CF ₃ COOH (300)	TBHP (3)	110	15
5	AlCl ₃ (100)	TBHP (3)	110	58
6	FeCl ₃ (100)	TBHP (3)	110	42
7	ZnCl ₂ (100)	TBHP (3)	110	<8
8	TiCl ₄ (100)	TBHP (3)	110	40
9	SnCl ₂ (100)	TBHP (3)	110	48
10	$Cu(OTf)_2$ (100)	TBHP (3)	110	38
11	AlCl ₃ (50)	TBHP (3)	110	57
12	AlCl ₃ (25)	TBHP (3)	110	56
13	AlCl ₃ (20)	TBHP (3)	110	51
14	AlCl ₃ (25)	TBHP (3)	95	65
15 ^c	AlCl ₃ (25)	TBHP (3)	95	71
16 ^c	AlCl ₃ (25)	TBHP (3.5)	95	72
17^c	AlCl ₃ (25)	TBHP (2)	95	51
18 ^c	AlCl ₃ (25)	aq TBHP(3)	95	47
19 ^c		TBHP (3)	95	<15

^aReaction conditions: 1 (0.25 mmol), **d** (1.25 mmol), time 18 h. ^bIsolated pure product. c Under N_{2} atmoshphere.

both weak (CH₃COOH) and strong (CF₃SO₃H) organic acids were used in lieu of CF₃COOH; however, poorer yields were obtained in both these cases (Table 1, entries 2 and 3). Even a 3-fold excess of CF₃COOH had no substantial effect on the product yield (Table 1, entry 4). These observations suggest that neither the strength of acid nor their concentrations are the determining factors for better conversion. Thus, we attempted the reaction in the presence of Lewis acids instead of protic acids.

Gratifyingly, the use of Lewis acid AlCl₃ afforded the acylated product in an improved yield of 58% (Table 1, entry 5). Encouraged by this result, other Lewis acids such as FeCl₃ (42%), ZnCl₂ (<8%), TiCl₄ (40%), SnCl₂ (48%), and Cu(OTf)₂ (38%) were tested, but all were found to be inferior to AlCl₃ (58%) (Table 1, entries 5–10). Interestingly, when the quantity of AlCl₃ was reduced to half (50 mol %) and quarter (25 mol %), the yield virtually remained unaltered (Table 1, entries 11 and 12). However, the yield dropped marginally when AlCl₃ loading was decreased to 20 mol % (Table 1, entry

13). A 9% increase in the yield was observed (Table 1, entry 14) upon performing the reaction at a lower temperature of 95 °C. The better yield (65%) obtained at lower temperature (95 °C) compared to the lower yield (56%) at higher temperature (110 °C) may be related to some of the thermodynamic parameters in the reaction as the acylated product formed is stable at high temperature. Maintaining the temperature at 95 °C and performing the reaction under N₂ atmosphere resulted in 6% further enhancement in the yield (Table 1, entry 15). No major improvement in the product yield was noticed with an increased amount of oxidant (3.5 equiv), while the use of 2 equiv was not sufficient for this transformation (Table 1, entries 16 and 17). Aqueous TBHP was found to be less effective compared to that of a decane solution of TBHP (Table 1, entry 18). Control experiment in the absence of Lewis acid AlCl₃ afforded acylated product in <15% suggesting the essential requirement of AlCl₃ to bring about the desired transformation (Table 1, entry 19). To check whether AlCl₃ has definite role or the acid (HCl) generated in the reaction medium is activating the N-atom a reaction was carried out in the presence of 1 equiv of HCl. The formation of product in a modest yield of 33% ascertains the distinct role of AlCl₃ in this transformation. From these screening studies, the optimal conditions established were the use of AlCl₃ (25 mol %) and TBHP (5-6 M in decane) (3 equiv) at 95 °C under N₂ atmosphere (Table 1, entry 15).

With these optimized conditions in hand, we examined the scope of these cross-dehydrogenative couplings by reacting isoquinoline (1) with a set of alkylbenzenes (a-i) possessing both electron-donating as well as electron-withdrawing substituents (Scheme 2). Under the present conditions, isoquinoline was smoothly acylated with various alkylbenzenes to afford the corresponding coupled products 1a-i in moderate to good yields as shown in Scheme 2. Substituents present in the phenyl ring of alkylbenzenes play a role in controlling the product yields as evident from their yields and the reaction times (Scheme 2). Methylbenzenes bearing additional -Me group(s) irrespective of their position like o-Me (b), m-Me (c), and p-Me (d) provided good yields of their corresponding coupled products (1b-d) with the retention of other -Me group(s), an observation consistent with our previous reports. 13a-f However, a slightly lower yield was obtained for ortho-substituted alkylbenzene; the difference in o-xylene (b) compared to meta and para analogues (1c and 1d) could be due to steric effects imparted by the ortho-substituent where a strongly electron-donating -OMe (e) group is expected to give a higher yield of the acylated product (1e, Scheme 2), but the actual result was contrary to that expected. This may be due to the coordination of the methoxy group of alkylbenzene (e) with the Lewis acid (AlCl₃), thereby making it less available for its catalytic activity. In order to expand the scope of this coupling reaction, isoquinoline substituted with various groups like 3-Me (2), 4-Br (3), and 5-NO₂ (4) were reacted with alkylbenzenes (a and d) (Scheme 2). As can be seen from Scheme 2, all of these isoquinolines underwent efficient coupling with alkylbenzenes (a and d) under the present reaction conditions to afford products 2a, 3a, 4a, and 4d in good to moderate yields (Scheme 2).

The next focus of this strategy was to acylate other electron-deficient heterocyclic compounds such as quinolines and quinoxalines (Scheme 3). Quinoline underwent cross-dehydrogenative coupling with different alkylbenzenes (a–i) to give exclusive C2-monoacylated products (5a–i, Scheme 3). It is to

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Scheme 2. Substrate Scope for C1 Acylation of Isoquinolines^{a,b}

^aReaction conditions: isoquinoline (0.5 mmol), alkylbenzenes (2.5 mmol), AlCl₃ (0.125 mmol), TBHP in decane (5–6 M) (3 equiv), 95 $^{\circ}$ C, time 18–24 h. ^bYields of the pure product reported.

be noted that in the case of quinolines the acylation took place regioselectively at its C2 position only with no traces of other regioisomers. However, in previous reports by Antonchick^{6h} and Prabhu¹² a mixture of both mono- (at the C2 position) and diacylated (at the C2 and C4 positions) products were formed in various proportions. No C4 aroylation product was observed even when the C2 position was blocked with a methyl or a tertbutyl group, suggesting the strong regioselective nature of the present transformation (Scheme 4). It is well-known that under acidic conditions (protic acid) both C-2 and C-4 positions are susceptible toward nucleophilic radical addition because of the N-protonation in quinoline. Due to the pronounced -I effect of the adjacent protonated N-atom there is a higher preference at the C-2 position. However, coordination of weak Lewis acid (AlCl₃) with the nitrogen of quinoline is just sufficient to activate the C-2 position and not the C-4, therefore giving only one regioisomer. The reactivity of different alkylbenzenes with quinoline is similar to that observed for their reaction with isoquinoline, but the overall yields of coupled products were marginally lower (Scheme 3). Other substituted quinolines like 8-methylquinoline (6) and 8-methoxyquinoline (7) reacted with alkylbenzenes (a, c, d, and f) to provide their respective acylated products (6a, 6c, 6d, 6f, and 7d) in moderate yields as shown in Scheme 3. Heterocycles bearing two N-atoms like quinoxaline (8) were also monofunctionalized in good yields (8a-h, Scheme 3). Similarly, electron-deficient heterocycle 7methylquinoxaline (9) when coupled with p-xylene (d) under

the optimized reaction conditions resulted in an inseparable mixture of two regioisomeric acylated products (9d, Scheme 3).

Next the mechanism of the direct acylation was investigated. Substantial quenching of product formation in the presence of radical inhibitor (TEMPO) was observed (Scheme 5). Along with the formation of a trace (~4%) of coupled product, a TEMPO ester (58%) was also isolated under the reaction conditions. This provides evidence for the formation of an acyl radical in the medium, thus supporting a radical pathway. The observed kinetic isotope effect $(k_{\rm H}/k_{\rm D}\sim1)$ (see the Supporting Information for calculations) during an intermolecular competing reaction of toluene and toluene- d_8 with isoquinoline implies that sp³ C–H bond cleavage is not the rate-determining step in this process (Scheme 6). On the basis of these observations and literature precedence, a plausible mechanism is proposed in Scheme 7.

In the presence of TBHP, alkylbenzene (a) is oxidized sequentially to benzyl alcohol (I) and benzaldehyde (II), which subsequently generates acyl radical (III) via the cleavage of aldehydic C–H bond. On the other hand, AlCl₃ coordinates with the *N*-atom of heterocycle 1 to form intermediate A, making the heterocyclic ring further electron-deficient. An acyl radical (III) formed in the medium is nucleophilic in nature and attacks at the more electrophilic C1 position of the heterocycle to form the corresponding radical intermediate B. Rearomatization of intermediate B provides the desired acylated product 1a (Scheme 7). When the reaction was

Scheme 3. Scope of N-Heterocyles^{a,b}

^aReaction conditions: 5-9 (0.5 mmol), alkylbenzenes (2.5 mmol), AlCl₃ (0.125 mmol), TBHP in decane (5-6 M) (3 equiv), 95 °C, time 18-24 h. ^bYields of the pure product reported.

Scheme 4. Demonstration of Regioselectivity in Quinoline

Scheme 5. Trapping of Acyl Intermediate with TEMPO

Scheme 6. Kinetic Isotope Study

performed either with benzyl alcohol or with benzaldehyde instead of toluene under otherwise identical conditions, the yields of product **1a** obtained were 84% and 90%, respectively, therefore supporting the intermediacy of benzyl alcohol and aldehyde in this reaction. The selective acylation could be explained in term of nucleophilic character of acyl radical **III** generated, which attacks at the more electrophilic position of coordinated heterocycle **A**. A similar mechanism is expected for other *N*-heterocycles.

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Scheme 7. Proposed Reaction Mechanism

CONCLUSION

In conclusion, we have developed an efficient, mild, and cost-effective method for the regiospecific acylation of electron-deficient N-heterocycles using methylbenzenes. In this transformation, Lewis acid $AlCl_3$ is used as catalyst and TBHP as oxidant. This reaction tolerates a wide range of functional groups and proceeds efficiently for the acylation of N-heterocycles. The reaction serves as complement to classical Minisci reaction.

■ EXPERIMENTAL SECTION

General Information. All the compounds were commercial grade and were used without further purification. Organic extracts were dried over anhydrous sodium sulfate. Solvents were removed in a rotary evaporator under reduced pressure. Silica gel (60–120 mesh size) was used for the column chromatography. Reactions were monitored by TLC on silica gel 60 F254 (0.25 mm). NMR spectra were recorded in CDCl₃ with tetramethylsilane as the internal standard for proton NMR (400 and 600 MHz) and with CDCl₃ solvent as internal standard for ¹³C NMR (100 and 150 MHz). HRMS spectra were recorded using ESI mode (Q-TOF type Mass Analyzer). IR spectra were recorded in KBr or neat.

General Procedure for the Synthesis of Isoquinolin-1-yl(p-tolyl)methanone (1d). Isoquinoline (0.5 mmol, 64.5 mg), AlCl₃ (0.125 mmol, 16.6 mg), p-xylene (2.5 mmol, 265 mg), and 5–6 M decane TBHP (1.5 mmol, 300 μ L) were added sequentially into an oven-dried 25 mL round-bottom flask. Then the resultant mixture was heated at 95 °C for 18 h under an atmosphere of N₂ sealed with a rubber septum. After completion of the reaction, as indicated by TLC, the reaction mixture was cooled to room temperature and admixed with ethyl acetate (30 mL). The organic layer was washed sequentially with a saturated solution of sodium bicarbonate (2 × 5 mL) and water (2 × 5 mL). The organic layer was dried over anhydrous Na₂SO₄ and evaporated under vacuum. The crude product so obtained was then purified by column chromatography using EtOAc and hexane (0.4:9.6) to give aroylated product 1d (88 mg, 71%).

Trapping of Radical Intermediates with Radical Scavenger TEMPO. Isoquinoline (0.5 mmol, 64.5 mg), AlCl₃ (0.125 mmol, 16.6 mg), toluene (2.5 mmol, 230 mg), and 5–6 M decane TBHP (1.5 mmol, 300 μ L) and TEMPO (1.5 mmol, 234 mg) were added sequentially into an oven-dried 25 mL round-bottom flask. Then the resultant mixture was heated at 95 °C for 18 h. After completion of the reaction, as indicated by TLC, the reaction mixture was cooled to room temperature and admixed with ethyl acetate (30 mL). The organic layer was washed sequentially with a saturated solution of sodium bicarbonate (2 × 5 mL) and water (2 × 5 mL). The organic layer was dried over anhydrous Na₂SO₄ and evaporated under vacuum. The crude products so obtained were then purified by column

chromatography using EtOAc and hexane as the eluents. TEMPO ester adduct 2,2,6,6-tetramethylpiperidin-1-yl benzoate (10a) was isolated in 58% yield along with a trace of 1a.

Kinetic Isotope Effect Studies. To a mixture of isoquinoline (1) (0.5 mmol, 64.5 mg) and AlCl₃ (0.125 mmol, 16.6 mg) was added an equimolar quantity of toluene (a) (1.25 mmol, 115 mg), and toluene d_8 (a') (1.25 mmol, 125 mg). To this resultant heterogeneous mixture was then added 5-6 M decane TBHP (300 µL, 1.5 mmol), and the resultant mixture was heated in a preheated oil bath at 95 °C for 18 h. The reaction mixture was admixed with water (10 mL), and the product was extracted with ethyl acetate (50 mL). The organic phase was dried over anhydrous sodium sulfate and concentrated in vacuo. The crude product so obtained was purified over a column of silica gel and eluted with hexane/ethyl acetate 9.4:0.4 to give the expected product in 42% yield (1a and 1a'). The ratio of the deuterated (1a') and nondeuterated (1a) aroylated product was calculated on the basis of the integration ratio of the aromatic proton peak at 7.48 (originating from toluene) and aromatic proton at 8.60 (originated from Nheterocycle) by adopting the procedure of Xie et al. Isoquinoline-1-yl(phenyl)methanone (1a):^{6h} orange gum; yield

Isoquinoline-1-yl(phenyl)methanone (1a):^{6h} orange gum; yield 72 mg, 62%; ¹H NMR (400 MHz, CDCl₃): δ (ppm) 7.47 (t, 2H, J = 7.6 Hz), 7.59–7.64 (m, 2H), 7.74 (t, 1H, J = 7.6 Hz), 7.81 (d, 1H, J = 6.0 Hz), 7.91–7.96 (m, 3H), 8.22 (d, 1H, J = 8.8 Hz), 8.60 (d, 1H, J = 5.6 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 122.8, 126.4, 126.6, 127.3, 128.5, 128.7, 130.9, 133.8, 136.8, 136.9, 141.4, 156.6, 194.9; IR (KBr) 3056, 2926, 2851, 1671, 1581, 1248, 1154, 1020 cm⁻¹; HRMS (ESI) calcd for C₁₆H₁₁NO (MH⁺) 234.0913, found 234.0919.

Isoquinolin-1-yl(o-tolyl)methanone (1b):^{6h} brown gum; yield 72 mg, 58%; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 2.53 (s, 3H), 7.21 (t, 1H, J = 7.8 Hz), 7.32 (d, 1H, J = 7.8 Hz), 7.40 (d, 1H, J = 7.8 Hz), 7.43 (t, 1H, J = 8.4 Hz), 7.67 (t, 1H, J = 6.6 Hz), 7.76 (t, 1H, J = 7.8 Hz), 7.79 (d, 1H, J = 6.0 Hz), 7.92 (d, 1H, J = 7.8 Hz), 8.62 (d, 1H, J = 8.4 Hz), 8.86 (d, 1H, J = 5.4 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 21.5, 123.1, 125.7, 126.5, 126.6, 127.3, 128.8, 130.9, 131.99, 132.0, 132.3, 137.0, 137.6, 139.9, 141.6, 157.2, 198.0; IR (KBr) 3057, 2926, 2855, 1669, 1576, 1243, 1156, 1041 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₃NO (MH⁺) 248.1070, found 248.1074.

Isoquinolin-1-yl(*m*-tolyl)methanone (1c):^{6h} orange gum; yield 84 mg, 68%; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.36 (s, 3H), 7.33 (t, 1H, J = 7.6 Hz), 7.40 (d, 1H, J = 7.6 Hz), 7.59 (t, 1H, J = 6.8 Hz), 7.71 (d, 2H, J = 9.6 Hz), 7.74 (d, 1H, J = 7.6 Hz), 7.78 (d, 1H, J = 8.6 Hz), 7.90 (d, 1H, J = 8.0 Hz), 8.49 (d, 1H, J = 8.4 Hz), 8.58 (d, 1H, J = 6.0 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 21.5, 122.7, 126.4, 126.5, 127.3, 128.3, 128.4, 128.6, 130.9, 131.2, 134.7, 136.8, 138.5, 141.4, 156.9, 195.2; IR (KBr) 3055, 2924, 2854, 1668, 1584, 1279, 1175, 1049 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₃NO (MH⁺) 248.1070, found 248.1076.

Isoquinolin-1-yl(*p*-tolyl)methanone (1d):. ^{6h,12} orange gum; yield 86 mg, 70%; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.42 (s, 3H), 7.27 (d, 2H, J = 8.0 Hz), 7.60 (t, 1H, J = 7.6 Hz), 7.73 (t, 1H, J = 6.8 Hz), 7.79 (d, 1H, J = 5.2 Hz), 7.85 (d, 2H, J = 8.4 Hz), 7.91 (d, 1H, J = 8.0 Hz), 8.91 (d, 1H, J = 8.4 Hz), 8.59 (d, 1H, J = 8.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 22.0, 122.6, 126.4, 126.6, 127.3, 128.4, 129.4, 130.9, 131.1, 134.3, 136.9, 141.4, 144.9, 157.0, 194.7; IR (KBr) 3054, 2923, 1665, 1604, 1249, 1152, 1018 cm⁻¹; HRMS (ESI) calcd for C_{1.7}H_{1.3}NO (MH⁺) 248.1070m found 248.1073.

Isoquinolin-1-yl(4-methoxyphenyl)methanone (1e):.^{6h,12} brown gum; yield 72 mg, 55%; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 3.84 (s, 3H), 6.92 (d, 2H, J = 9.0 Hz), 7.57 (t, 1H, J = 7.8 Hz), 7.70 (t, 1H, J = 7.8 Hz), 7.76 (d, 1H, J = 5.4 Hz), 7.88 (d, 1H, J = 8.4 Hz), 7.91 (d, 2H, J = 8.4 Hz), 8.14 (d, 1H, J = 9.0 Hz), 8.56 (d, 1H, J = 6.0 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 55.7, 114.0, 122.4, 126.5, 127.2, 128.3, 129.8, 130.0, 130.9, 133.3, 136.9, 141.4, 157.3, 164.3, 193.6; IR (KBr) 3055, 2932, 2840, 1661, 1596, 1422, 1254, 1171, 1074 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₃NO₂ (MH⁺) 264.1019, found 264.1017.

(4-Chlorophenyl)(isoquinolin-1-yl)methanone (1f): ^{14b} brown solid; yield 76 mg, 57%; mp 73–75 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.43 (d, 2H, J = 8.4 Hz), 7.62 (t, 1H, J = 8.0 Hz), 7.74 (t, 1H, J = 8.0 Hz), 7.80 (d, 1H, J = 5.2 Hz), 7.89–7.92 (m, 3H), 8.23 (d, 1H, J = 8.0 Hz), 7.80 (d, 1H, J = 8.0 Hz), J

J = 8.4 Hz), 8.58 (d, 1H, J = 5.6 Hz); 13 C NMR (100 MHz, CDCl₃) δ (ppm) 123.2, 126.3, 126.7, 127.4, 128.7, 129.0, 131.0, 132.4, 135.3, 137.0, 140.4, 141.3, 155.9, 193.6; IR (KBr) 3054, 2926, 2854, 1664, 1584, 1249, 1090 cm⁻¹; HRMS (ESI) calcd for $C_{16}H_{10}CINO$ (MH⁺) 268.0524, found 268.0528.

(4-Bromophenyl)(isoquinolin-1-yl)methanone (1g):¹² brown solid; yield 91 mg, 59%; mp 70–72 °C; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 7.60–7.66 (m, 3H), 7.75 (t, 1H, J = 8.0 Hz), 7.82–7.85 (m, 3H), 7.93 (d, 1H, J = 8.4 Hz), 8.26 (d, 1H, J = 8.8 Hz), 8.59 (d, 1H, J = 8.4 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 123.2, 126.3, 126.7, 127.4, 128.7, 129.2, 131.0, 132.0, 132.4, 135.7, 137.0, 141.3, 155.8, 193.8; IR (KBr) 3053, 2925, 2854, 1662, 1580, 1393, 1247, 1070 cm $^{-1}$; HRMS (ESI) calcd for $C_{16}H_{10}B$ rNO (MH $^+$) 312.0019, found 312.0024.

(3,5-Dimethylphenyl)(isoquinolin-1-yl)methanone (1h):^{6h} yellow gum; yield 92 mg, 70%; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.33 (s, 6H), 7.24 (s, 1H), 7.54 (s, 2H), 7.60 (t, 1H, J = 8.4 Hz), 7.73 (t, 1H, J = 6.8 Hz), 7.79 (d, 1H, J = 5.6 Hz), 7.91 (d, 1H, J = 8.4 Hz), 8.17 (d, 1H, J = 8.4 Hz), 8.59 (d, 1H, J = 5.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 21.3, 122.6, 126.4, 126.5, 127.2, 128.4, 128.6, 130.8, 135.7, 136.8, 136.9, 138.3, 141.3, 157.1, 195.5; IR (KBr) 3054, 2919, 2859, 1668, 1601, 1453, 1299, 1144, 1085 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₅NO (MH⁺) 262.1226, found 262.1233.

(3-lodo-5-methylphenyl)(isoquinolin-1-yl)methanone (1i): brown gum; yield 111 mg, 59%; $^1\mathrm{H}$ NMR (400 MHz, CDCl₃) δ (ppm) 2.36 (s, 3H), 7.65 (t, 1H, J = 8.0 Hz), 7.70 (s, 1H), 7.74–7.78 (m, 2H), 7.84 (d, 1H, J = 5.6 Hz), 7.94 (d, 1H, J = 8.0 Hz), 8.06 (s, 1H), 8.23 (d, 1H, J = 8.4), 8.60 (d, 1H, J = 6.0 Hz); $^{13}\mathrm{C}$ NMR (100 MHz, CDCl₃) δ (ppm) 21.1, 94.3, 123.2, 126.3, 126.7, 128.8, 130.8, 131.0, 135.3, 136.8, 138.7, 140.6, 14.6, 141.4, 143.1, 155.8, 193.6; IR (KBr) 2923, 2853, 1667, 1562, 1429, 1277, 1146, 1018 cm $^{-1}$; HRMS (ESI) calcd for $\mathrm{C}_{17}\mathrm{H}_{12}\mathrm{INO}$ (MH $^+$) 374.0036, found 374.0036.

(3-Methylisoquinolin-1-yl)(phenyl)methanone (2a): brown solid; yield 64 mg, 52%; mp 95–97 °C; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 2.71 (s, 3H), 7.38–7.49 (m, 3H), 7.57–7.60 (m, 2H), 7.65 (t, 1H, J = 9.6 Hz), 7.79 (d, 1H, J = 7.8 Hz), 7.95 (d, 2H, J = 6.6 Hz), 8.05 (d, 1H, J = 7.8 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 24.3, 120.6, 124.6, 126.1, 126.4, 126.7, 127.3, 128.6, 130.7, 131.0, 133.9, 136.6, 137.5, 150.4, 195.1; IR (KBr) 3059, 2924, 2855, 1670, 1589, 1447, 1239, 1027 cm⁻¹; HRMS (ESI) calcd for C_{17} H13NO (MH⁺) 248.1070, found 248.1072.

(4-Bromoisoquinolin-1-yl)(phenyl)methanone (3a): red solid; yield 101 mg, 65%; mp 129–130 °C; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 7.48 (t, 2H, J = 7.8 Hz), 7.62 (t, 1H, J = 7.2 Hz), 7.69 (t, 1H, J = 7.8 Hz), 7.87 (t, 1H, J = 8.4 Hz), 7.94 (d, 2H, J = 7.8 Hz), 8.23 (d, 1H, J = 9.0 Hz), 8.29 (d, 1H, J = 9.0 Hz), 8.80 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 122.0, 126.7, 126.9, 127.8, 128.7, 129.4, 130.9, 132.2, 134.1, 135.7, 136.6, 143.1, 155.9, 194.2; IR (KBr) 3056, 2961, 2855, 1659, 1523, 1268, 1158, 1018 cm⁻¹; HRMS (ESI) calcd for C₁₆H₁₀BrNO (MH⁺) 312.0019, found 312.0025.

(5-Nitroisoquinolin-1-yl)(phenyl)methanone (4a): white solid; yield 92 mg, 66%; mp 106–108 °C; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 7.49 (t, 2H, J = 7.8 Hz), 7.64 (t, 1H, J = 7.2 Hz), 7.72 (t, 1H, J = 7.8 Hz), 7.93 (d, 2H, J = 7.8 Hz) 8.55 (t, 2H, J = 7.2 Hz), 8.62 (d, 1H, J = 6.0 Hz), 8.82 (d, 1H, J = 6.6 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 117.8, 126.89, 126.92, 128.7, 128.8, 129.3, 131.0, 133.4, 134.4, 136.2, 144.6, 145.5, 157.3, 193.9; IR (KBr) 3066, 2922, 2844, 1661, 1526, 1353, 1276, 1161, 1069 cm⁻¹; HRMS (ESI) calcd for C₁₆H₁₀N₂O₃ (MH⁺) 279.0764, found 279.0770.

(5-Nitroisoquinolin-1-yl)(p-tolyl)methanone (4d): brown solid; yield 98 mg, 67%; mp 107–109 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.44 (s, 3H), 7.30 (d, 2H, J = 8.4 Hz), 7.72 (t, 1H, J = 8.4 Hz), 7.83 (d, 2H, J = 8.0 Hz), 7.50–8.57 (m, 2H), 8.62 (d, 1H, J = 6.4 Hz), 8.81 (t, 1H, J = 6.4 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 22.0, 117.6, 119.5, 126.8, 126.9, 128.7, 129.3, 129.6, 131.1, 133.5, 133.8, 134.0, 144.7, 157.7, 193.6; IR (KBr) 3063, 2924, 2853, 1669, 1448, 1246, 1215, 1180, 1024 cm⁻¹; HRMS (ESI) calcd for $C_{17}H_{12}N_2O_3$ (MH⁺) 293.0921, found 293.0922.

Phenyl(quinolin-2-yl)methanone (5a): ^{14a} red solid; yield 68 mg, 58%; mp 96–98 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.50

(t, 2H, J=7.6 Hz), 7.59–7.66 (m, 2H), 7.77 (t, 1H, J=7.2 Hz), 7.89 (d, 1H, J=8.4 Hz), 8.09 (d, 1H, J=8.4 Hz), 8.17–8.22 (m, 3H), 8.33 (d, 1H, J=8.4 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 121.0, 127.8, 128.4, 128.6, 129.1, 130.3, 130.8, 131.6, 133.3, 136.4, 137.3, 147.0, 154.9, 194.0; IR (KBr) 3054, 2925, 2853, 1661, 1450, 1317, 1168, 1019 cm⁻¹; HRMS (ESI) calcd for $C_{16}H_{11}$ NO (MH⁺) 234.0913, found 234.0915.

Quinolin-2-yl(o-tolyl)methanone (5b): yellow solid; yield 67 mg, 54%; mp 79–80 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.44 (s, 3H), 7.28 (t, 1H, J = 7.6 Hz), 7.33 (d, 1H, J = 7.6 Hz), 7.45 (t, 1H, J = 7.2 Hz), 7.60 (d, 1H, J = 8.0 Hz), 7.65 (t, 1H, J = 7.2 Hz), 7.75 (t, 1H, J = 8.0 Hz), 7.90 (d, 1H, J = 8.0 Hz), 8.14 (t, 2H, J = 8.4 Hz), 8.34 (d, 1H, J = 8.0 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 21.0, 120.5, 125.2, 127.8, 128.8, 129.2, 130.2, 131.0, 131.3, 131.4, 137.3, 137.4, 138.8, 147.3, 155.0, 197.7; IR (KBr) 3055, 2923, 2853, 1660, 1561, 1454, 1313, 1260, 1092 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₃NO (MH⁺) 248.1070, found 248.1079.

Quinolin-2-yl(m-tolyl)methanone (5c): white solid; yield 77 mg, 62%; mp 80–82 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.43 (s, 3H), 7.38 (d, 1H, J = 8.0 Hz), 7.43 (t, 1H, J = 7.2 Hz), 7.66 (t, 1H, J = 7.2 Hz), 7.79 (t, 1H, J = 7.2 Hz), 7.91 (d, 1H, J = 8.0 Hz), 8.00 (d, 2H, J = 6.8 Hz), 8.08 (d, 1H, J = 8.4 Hz), 8.21 (d, 1H, J = 8.4 Hz), 8.34 (d, 1H, J = 8.8 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 21.6, 121.0, 127.8, 128.2, 128.5, 129.0, 129.1, 130.3, 130.8, 131.9,134.1, 136.3, 137.2, 138.1, 146.9, 155.1, 194.4; IR (KBr) 3052, 2924, 2856, 1660, 1596, 1458, 1313, 1139, 1139, 1013 cm⁻¹; HRMS (ESI) calcdfor C₁₇H₁₃NO (MH⁺) 248.1070, found 248.1067.

Quinolin-2-yl(*p*-tolyl)methanone (5d):¹² brown solid; yield 78 mg, 63%; mp 62–64 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.44 (s, 3H), 7.30 (d, 2H, J = 8.0 Hz), 7.64 (t, 1H, J = 6.8 Hz), 7.76 (t, 1H, J = 7.6 Hz), 7.88 (d, 1H, J = 8.0 Hz), 8.06 (d, 1H, J = 8.4 Hz), 8.13 (d, 2H, J = 8.0 Hz), 8.18 (d,1H, J = 8.4 Hz), 8.32 (d, 1H, J = 8.8 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 22.0, 121.0, 127.9, 128.5, 129.07, 129.13, 130.3, 130.7, 131.8, 133.8, 137.2, 144.2, 147.0, 155.3, 193.7; IR (KBr) 3066, 2922, 2852, 1659, 1456, 1317, 1161, 1112 cm⁻¹; HRMS (ESI) calcdfor C₁₇H₁₃NO (MH⁺) 248.1070, found 248.1077.

(4-Methoxyphenyl)(quinolin-2-yl)methanone (5e): 14b white solid; yield 65 mg, 49%; mp 65–68 °C; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 3.87 (s, 3H), 6.99 (d, 2H, J = 7.2 Hz), 7.65 (t, 1H, J = 7.6 Hz), 7.78 (t, 1H, J = 7.2 Hz), 7.90 (d, 1H, J = 7.6 Hz), 8.06 (d, 1H, J = 8.4 Hz), 8.20 (d, 1H, J = 8.0 Hz), 8.28 (d, 2H, J = 6.8 Hz), 8.33 (d, 1H, J = 8.4 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 55.7, 113.8, 121.1, 127.9, 128.4, 129.0, 129.2, 130.2, 130.7, 134.1, 137.2, 146.9, 155.6, 163.9, 192.4; IR (KBr) 3059, 2927, 2848, 1652, 1507, 1319, 1256, 1157, 1025 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₃NO₂ (MH⁺) 264.1019, found 264.1025.

(4-Chlorophenyl)(quinolin-2-yl)methanone (5f): brown solid; yield 68 mg, 51%; mp 105-107 °C; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 7.47 (d, 2H, J = 8.4 Hz), 7.65 (t, 1H, J = 8.4 Hz), 7.78 (t, 1H, J = 6.8 Hz), 7.89 (d, 1H, J = 7.6 Hz), 8.11 (d, 1H, J = 8.8 Hz), 8.17 (d, 1H, J = 8.4 Hz), 8.22 (d, 2H, J = 8.8 Hz), 8.34 (d, 1H, J = 8.8 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 120.9, 127.9, 128.7, 128.8, 129.2, 130.4, 130.7, 133.1, 134.8, 137.5, 139.8, 146.9, 154.5, 192.6; IR (KBr) 2926, 2854, 1663, 1590, 1459, 1316, 1209, 1170, 1089 HRMS (ESI) calcd for C₁₆H₁₀CINO (MH $^+$) 268.0524, found 268.0521.

(4-Bromophenyl)(quinolin-2-yl)methanone (5g): brown solid; yield 84 mg, 54%; mp 111–113 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.65 (t, 3H, J = 9.6 Hz), 7.78 (t, 1H, J = 9.2 Hz), 7.89 (d, 1H, J = 7.6 Hz), 8.10–8.17 (m, 4H), 8.33 (d, 1H, J = 8.4 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 120.9, 127.9, 128.5, 128.8, 129.2, 130.4, 130.7, 131.6, 133.2, 135.2, 137.4, 146.9, 154.4, 192.7; IR (KBr) 3048, 2956, 2926, 1664, 1586, 1458, 1316, 1167, 1068 cm $^{-1}$; HRMS (ESI) calcd for C₁₆H₁₀BrNO (MH $^+$) 312.0019, found 312.0026.

(3,5-Dimethylphenyl)(quinolin-2-yl)methanone (5h): red solid; yield 84 mg, 64%; mp 87–89 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.37 (s, 6H), 7.24 (s, 1H), 7.64 (t, 1H, J = 8.0 Hz), 7.75–7.79 (m, 3H), 7.89 (d, 1H, J = 8.0 Hz), 8.03 (d, 1H, J = 8.8 Hz), 8.19 (d, 1H, J = 8.4 Hz), 8.32 (d, 1H, J = 8.8 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 21.5, 121.0, 127.8, 128.5, 129.1, 129.3, 130.2, 130.8, 135.1, 136.4, 137.2, 138.0, 147.0, 155.4, 194.7; IR (KBr) 3060,

2923, 2855, 1662, 1598, 1461, 1325, 1143, 1035 $cm^{-1};$ HRMS (ESI) calcd for $C_{18}H_{15}NO\ (MH^+)$ 262.1226, found 262.1229.

(3-lodo-5-methylphenyl)(quinolin-2-yl)methanone (5i): brown solid; yield 106 mg, 57%; mp 121–123 °C; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 2.37 (s, 3H), 7.66 (t, 1H, J = 7.2 Hz), 7.76–7.80 (m, 2H), 7.89 (d, 1H, J = 8.0 Hz), 7.95 (s, 1H), 8.08 (d, 1H, J = 8.8 Hz), 8.19 (d, 1H, J = 8.0 Hz), 8.33 (d, 2H, J = 8.8 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 21.2, 93.9, 120.9, 127.9, 128.8, 129.2, 130.4, 130.8, 131.4, 137.4, 137.6, 138.1, 140.2, 142.5, 147.0, 154.3, 192.6; IR (KBr) 2924, 2853, 1664, 1563, 1311, 1143, 1016 cm $^{-1}$; HRMS (ESI) calcd for $C_{17}H_{12}$ INO (MH $^+$) 374.0036, found 374.0041.

(8-Methylquinolin-2-yl)(phenyl)methanone (6a): ^{14c} white solid; yield 70 mg, 57%; mp 63–64 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 2.78 (s, 3H), 7.50–7.57 (m, 3H), 7.63 (t, 2H, J = 7.2 Hz), 7.74 (d, 1H, J = 8.0 Hz), 8.19 (d, 1H, J = 8.8 Hz), 8.31 (d, 1H, J = 8.4 Hz), 8.36 (d, 2H, J = 8.8 Hz); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 18.1, 120.6, 125.7, 128.1, 128.6, 129.2, 130.3, 131.9, 132.9, 136.8, 137.4, 139.1, 146.1, 153.3, 193.7; IR (KBr) 3056, 2953, 2922, 1660, 1594, 1445, 1322, 1286, 1159, 1070 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₃NO (MH⁺) 248.1070, found 248.1078.

(8-Methylquinolin-2-yl)(*m*-tolyl)methanone (6c): yellow liquid; yield 77 mg, 59%; 1 H NMR (600 MHz, CDCl₃) δ (ppm) 2.44 (s, 3H), 2.77 (s, 3H), 7.38 (t, 1H, J = 7.8 Hz), 7.42 (d, 1H, J = 7.8 Hz), 7.52 (t, 1H, J = 7.8 Hz), 7.62 (d, 1H, J = 6.6 Hz), 7.71 (d, 1H, J = 8.4 Hz), 8.15 (t, 2H, J = 8.4 Hz), 8.20 (s, 1H), 8.28 (d, 1H, J = 9.0 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 18.1, 21.6, 120.6, 125.7, 127.9, 128.5, 129.15, 129.17, 130.2, 132.4, 133.7, 136.7, 137.3, 137.7, 139.0, 146.0, 153.4, 193.7; IR (KBr) 3043, 2955, 2923, 1659, 1584, 1465, 1374, 1287, 1141, 1079 cm $^{-1}$; HRMS (ESI) calcd for C $_{18}$ H $_{15}$ NO (MH $^+$) 262.1226, found 262.1228.

(8-Methylquinolin-2-yl)(p-tolyl)methanone (6d): yellow liquid; yield 81 mg, 62%; 1 H NMR (400 MHz, CDCl $_3$) δ (ppm) 2.47 (s, 3H), 2.80 (s, 3H), 7.32 (d, 2H, J = 8.0 Hz), 7.54 (t, 1H, J = 8.0 Hz), 7.63 (d, 1H, J = 7.2 Hz), 7.73 (d, 1H, J = 8.0 Hz), 8.16 (d, 1H, J = 8.4 Hz), 8.28–8.31 (m, 3H); 13 C NMR (150 MHz, CDCl $_3$) δ (ppm) 18.2, 22.0, 120.7, 125.7, 128.4, 128.9, 129.2, 130.2, 132.1, 134.2, 137.4, 139.0, 143.8, 146.1, 153.7, 193.2; IR (KBr) 3093, 2922, 1653, 1463, 1319, 1250, 1156, 1020 cm $^{-1}$; HRMS (ESI) calcd for $C_{18}H_{15}NO$ (MH $^+$) 262.1226, found 262.1219.

(4-Chlorophenyl)(8-methylquinolin-2-yl)methanone (6f): yellow solid; yield 67 mg, 48%; mp 79–81 °C; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 2.78 (s, 3H), 7.50 (d, 2H, J = 8.8 Hz), 7.56 (t, 1H, J = 7.6 Hz), 7.65 (t, 1H, J = 7.6 Hz), 7.74 (d, 1H, J = 8.4 Hz), 8.19 (d, 1H, J = 8.4 Hz), 8.32 (d, 1H, J = 8.0 Hz), 8.35 (d, 2H, J = 8.4 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 18.2, 120.6, 125.8, 128.4, 128.8, 129.3, 130.4, 133.4, 135.2, 137.6, 139.0, 139.4, 146.0, 152.9, 192.3; IR (KBr) 2962, 2923, 2855, 1638, 1459, 1311, 1089 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₂ClNO (MH⁺) 282.0680, found 282.0685.

(8-Methoxyquinolin-2-yl)(*p***-tolyl)methanone (7d):** red liquid; yield 64 mg, 46%; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 2.43 (s, 3H), 4.05 (s, 3H), 7.10 (d, 1H, J = 7.2 Hz), 7.30 (d, 2H, J = 8.4 Hz), 7.44 (d, 1H, J = 7.2 Hz), 7.56 (t, 1H, J = 8.4 Hz), 8.09 (d, 1H, J = 8.8 Hz), 8.24–8.29 (m, 3H); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 21.9, 56.4, 108.6, 119.5, 121.6, 127.2, 128.9, 130.2, 132.1, 133.8, 137.0, 138.9, 144.0, 153.9, 156.5, 193.0; IR (KBr) 3066, 2928, 2838, 1655, 1606, 1465, 1325, 1260, 1131, 1040 cm $^{-1}$; HRMS (ESI) calcd for C₁₈H₁₅NO₂ (MH $^{+}$) 278.1176, found 278.1172.

Phenyl(quinoxalin-2-yl)methanone (8a): ^{14a} black solid; yield 86 mg, 74%; mp 100–102 °C; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 7.52 (t, 2H, J = 7.8 Hz), 7.64 (t, 1H, J = 7.8 Hz), 7.84 (t, 1H, J = 6.6 Hz), 7.88 (t, 1H, J = 8.4 Hz), 8.19 (m, 2H), 8.22 (d, 2H, J = 7.8 Hz), 9.47 (s, 1H); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 128.6, 129.6, 130.6, 130.9, 131.0, 131.5, 132.2, 132.4, 133.8, 143.3, 143.4, 145.5, 192.5; IR (KBr) 3062, 2925, 2853, 1659, 1597, 1322, 1234, 1127, 1014 cm⁻¹; HRMS (ESI) calcd for C₁₅H₁₀N₂O (MH⁺) 235.0866, found 235.0869.

Quinoxalin-2-yl(*m***-tolyl)methanone (8c):** brown gum; yield 92 mg, 74%; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 2.44 (s, 3H), 7.43 (t, 1H, J = 8.0 Hz), 7.48 (d, 1H, J = 7.2 Hz), 7.85–7.93 (m, 2H), 8.00 (s, 2H), 8.22 (d, 2H, J = 8.8 Hz), 9.47 (s, 1H); 13 C NMR (150 MHz,

CDCl₃) δ (ppm) 21.6, 128.5, 128.9, 129.6, 130.7, 130.9, 131.7, 132.2, 134.7, 135.8, 138.5, 140.7, 143.3, 145.5, 149.1, 192.9; IR (KBr) 2924, 2854, 1658, 1489, 1317, 1151, 1017 cm⁻¹; HRMS (ESI) calcd for $C_{16}H_{12}N_2O$ (MH⁺) 249.1022, found 249.1028.

Quinoxalin-2-yl(p-tolyl)methanone (8d):.^{6h,12,14a} red solid; yield 98 mg, 79%; mp 97–99 °C; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 2.42 (s, 3H), 7.30 (d, 2H, J = 7.6 Hz), 7.78–7.87 (m, 2H), 8.11 (d, 2H, J = 8.4 Hz), 8.16 (d, 2H, J = 8.4 Hz), 9.42 (s, 1H); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 22.0, 129.3, 129.5, 130.5, 130.9, 131.5, 132.0, 133.1, 140.6, 143.2, 144.8, 145.5, 149.1, 192.0; IR (KBr) 3053, 2922, 2851, 1657, 1601, 1324, 1229, 1157, 1121 cm $^{-1}$; HRMS (ESI) calcd for C₁₆H₁₂N₂O (MH $^{+}$) 249.1022, found 249.1019.

(4-Chlorophenyl)(quinoxalin-2-yl)methanone (8f): 14d brown solid; yield 90 mg, 67%; mp 85–87 °C; 1 H NMR (600 MHz, CDCl₃) δ (ppm) 7.50 (d, 2H, J = 8.4 Hz), 7.83–7.88 (m, 1H), 7.90 (t, 1H, J = 8.4 Hz), 8.17–8.23 (m, 4H), 9.49 (s, 1H); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 129.0, 129.7, 130.7, 130.9, 131.2, 132.5, 132.9, 134.1, 140.5, 143.3, 145.5, 148.4, 191.2; IR (KBr) 2926, 2855, 1663, 1460, 1313, 1164, 1087 cm $^{-1}$; HRMS (ESI) calcd for $C_{15}H_{9}ClN_{2}O$ (MH $^{+}$) 269.0476, found 269.0480.

(3,5-Dimethylphenyl)(quinoxalin-2-yl)methanone (8h): red solid; yield 102 mg, 78%; mp 151–152 °C; ¹H NMR (600 MHz, CDCl₃) δ (ppm) 2,40 (s, 6H), 7.30 (s, 1H), 7.78 (s, 2H), 7.86–7.91 (m, 2H), 8.20–8.22 (m, 2H), 9.43 (s, 1H); ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 21.5, 129.1, 129.6, 130.7, 131.0, 132.1, 135.7, 135.8, 138.3, 140.8, 143.3, 145.4, 149.4, 193.2; IR (KBr) 3063, 2920, 2859, 1659, 1601, 1327, 1214, 1149, 1031 cm⁻¹ HRMS (ESI) calcd for C₁₇H₁₄N₂O (MH⁺) 263.1179, found 263.1184.

(6-Methylquinoxalin-2-yl)(*p*-tolyl)methanone and (7-methylquinoxalin-2-yl)(*p*-tolyl)methanone (9d)L. red semisolid; yield 95 mg, 72%; 1 H NMR (400 MHz, CDCl₃) δ (ppm) 2.44 (s, 3H), 2.62 (s, 3H), 7.31 (d, 2H, J = 8.0 Hz), 7.65–7.71 (m, 1H), 7.94 (s, 1H), 8.06 (d, 1H, J = 8.4 Hz), 8.11 (d, 2H, J = 8.0 Hz), 9.39 (d, 1H, J = 15.6 Hz); 13 C NMR (150 MHz, CDCl₃) δ (ppm) 22.0, 22.3, 22.9, 128.4, 129.1, 129.3, 129.4, 130.2, 131.6, 133.31, 133.34, 134.5, 139.2, 140.8, 141.7, 141.9, 143.2, 143.4, 144.7, 144.75, 144.8, 145.6, 148.4, 149.2, 192.25, 192.31; IR (KBr) 2953, 2924, 2854, 1656, 1497, 1322, 1276, 1167, 1017 cm $^{-1}$; HRMS (ESI) calcd for C $_{17}$ H $_{14}$ N $_{2}$ O (MH $^+$) 263.1179, found 263.1174.

2,2,6,6-Tetramethylpiperidin-1-yl benzoate (10a): ^{13f} yellow gum; yield 76 mg, 58%; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 1.16 (s, 6H), 1.31 (s, 6H), 1.48–1.52 (broad singlet, 1H), 1.61–1.64 (broad singlet, 2H), 1.71 (m, 3H), 7.50 (t, 2H, J = 7.6 Hz), 7.61 (t, 1H, J = 6.8 Hz), 8.11 (d, 2H, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 17.1, 21.0, 32.1, 39.2, 60.5, 128.6, 129.7, 133.0, 166.5; IR (KBr) 2974, 2934, 1749, 1540, 1378, 1255, 1176, 1081, cm⁻¹; HRMS (ESI) calcd for C₁₆H₂₃NO₂ (MH⁺) 262.1802, found 262.1806.

ASSOCIATED CONTENT

S Supporting Information

Kinetic isotope effect experiment and spectral data for all compounds. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b00501.

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Notes

The authors declare no competing financial interest.

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